Optimisation of Glucose Concentration in Mixotrophic Cultivations of a Lichen Phycobiont *Ellipitochloris subphaerica*

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1. Introduction

The transition towards utilizing renewable resources for bioenergy has sparked considerable interest, with a particular focus on microalgae known for their high lipid content. Among microalgae, *Botryococcus braunii*, a trebouxiophycean green alga, is one of such microalgae and known for its high production of extracellular long-chain hydrocarbons, comprising up to 86% of its total dry weight (Borowitzka, 2018). However, *B. braunii* grow very slowly in culture, which raises the cost of hydrocarbon production.

Elliptochloris subsphaerica, a close relative of *B. braunii*, is a phycobiont of a lichen. In a previous study conducted at the laboratory to which I am affiliated, *E. subsphaerica* exhibited fast growth (3.5 g L^{-1} DCW in 14 days) and significant oil accumulation (30% of DCW) when cultured in medium X with 2% glucose (Shen, 2021). However, *E. subsphaerica* did not thrive in GTY medium, also containing 2% glucose (Shen, 2021), prompting questions about the role of glucose in its cultivation.

This experiment aims to identify the best glucose concentration for cultivating *E. subphaerica* with high biomass and lipid ratio. The effects of glucose concentrations in medium X ranging from 0% to 4% on growth characteristics and lipid accumulation were assessed.

2. Material & Methods

<u>2.1. Culture media & Conditions</u>: Five medium X with glucose concentrations of 0, 10, 20, 30, & 40 g L^{-1} each, along with a constant rate of other substances were prepared. Cells were inoculated at a concentration of $1x10^6$ cells per *mL*, under the light intensity at $250\mu \mod m^{-2} s^{-1}$ at $21^{\circ c}$, 100 rpm.

<u>2.2. Cell Growth</u>: The cellular growth was monitored biweekly, with observations taken once every two days via manual cell count by C-Chip (F01). Dry Cell Weight (DCW) was measured at the start and end of each experiment. Cells were harvested by centrifugation, froze at -80°C, and freeze-dried overnight.

2.3. Statistical Analysis: The analysis of data was carried out by Prism 9 software. The statistical significance was determined by analysis of variance (ANOVA). A P value < 0.05 was considered as statistically significant.

<u>2.4. Observation of Lipid Accumulation</u>: Cells collected on day 14 were stained with Nile-red dye at a ratio of 1:100 and observed under a fluorescent microscope.

3. Results & Discussion

<u>3.1. Growth Characteristics</u>: *E. subphaerica* did not present any significant differences in growth rate among the conditions, Fig 1 (p = 0.8912 - 0.9941). Mixotrophically cultured green algae

Chlorella such as pyrenoidosa has been known utilise to monosaccharides such glucose for their as growth (Zhang el. al., 2014). On the contrary, my result indicates that



E. subphaerica is not able to use glucose in mixotrophic condition but utilises alternative substances as its carbon source in the medium X, which differs from many previous reports. Moreover, *E. subsphaerica* achieved a high biomass yield (2.55 - 3.94 g L^{-1} DCW) and cell density (27.2 million cells mL¹) over two weeks using a glucose-free medium. This suggests potential cost reduction in large-scale cultivation of *E. subsphaerica* for bioenergy, reducing the dependence on glucose.

3.2. Lipid Analysis:

Lipid droplets (LD) were observed in all conditions, with no significant difference (p > 0.9999) related to growth rate. Optimising LD accumulation and identifying unknown lipid constituents are crucial for utilizing *E. subphaerica* as a viable resource for oil production. If the significant portion of the unidentified accumulated lipid (approximately 80%) hydrocarbons (Shen, 2021), it would further enhance its potential as a valuable source for hydrocarbon production.

References

Borowitzka, M.A. (2018). Biology of microalgae. 23–72 Zhang et al. (2014). Bioresource Technology. 173, 52–58 Shen. Y. (2021). Tsukuba Journal of Biology.